


FORM PTO-1390 (Modified) (REV 5-93)		U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE		ATTORNEY'S DOCKET NUMBER	
TRANSMITTAL LETTER TO THE UNITED STATES DESIGNATED/ELECTED OFFICE (DO/EO/US) CONCERNING A FILING UNDER 35 U.S.C. 371				065691/0209 09/762248	
		U.S. APPLICATION NO. (If known, see 37 C.F.R. 1.5) Unassigned			
INTERNATIONAL APPLICATION NO. PCT/FR99/01828		INTERNATIONAL FILING DATE July 26, 1999		PRIORITY DATE CLAIMED August 5, 1998	
TITLE OF INVENTION USE OF AN ANIMAL MODEL DEFICIENT IN P53 AND HAVING MEMORY DEFICIENCY AND/OR BEHAVIOURAL DISORDERS FOR THERAPEUTIC PURPOSES					
APPLICANT(S) FOR DO/EO/US Robert AMSON, Jean-Michel LASSALLE and Adam TELERMAN					
Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:					
1.	<input checked="" type="checkbox"/>	This is a FIRST submission of items concerning a filing under 35 U.S.C. 371.			
2.	<input type="checkbox"/>	This is a SECOND or SUBSEQUENT submission of items concerning a filing under 35 U.S.C. 371.			
3.	<input type="checkbox"/>	This express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1).			
4.	<input checked="" type="checkbox"/>	A proper Demand for International Preliminary Examination was made by the 19 th month from the earliest claimed priority date.			
5.	<input checked="" type="checkbox"/>	A copy of the International Application as filed (35 U.S.C. 371(c)(2))			
	<input type="checkbox"/>	is transmitted herewith (required only if not transmitted by the International Bureau).			
	<input checked="" type="checkbox"/>	has been transmitted by the International Bureau.			
	<input type="checkbox"/>	is not required, as the application was filed in the United States Receiving Office (RO/US)			
6.	<input checked="" type="checkbox"/>	A translation of the International Application into English (35 U.S.C. 371(c)(2)).			
7.	<input checked="" type="checkbox"/>	Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3))			
	<input type="checkbox"/>	are transmitted herewith (required only if not transmitted by the International Bureau).			
	<input type="checkbox"/>	have been transmitted by the International Bureau.			
	<input type="checkbox"/>	have not been made; however, the time limit for making such amendments has NOT expired.			
	<input checked="" type="checkbox"/>	have not been made and will not be made.			
8.	<input type="checkbox"/>	A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).			
9.	<input type="checkbox"/>	An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)).			
10.	<input type="checkbox"/>	A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).			
11.	<input type="checkbox"/>	Applicant claims small entity status under 37 CFR 1.27.			
Items 12. to 17. below concern other document(s) or information included:					
12.	<input type="checkbox"/>	An Information Disclosure Statement under 37 CFR 1.97 and 1.98.			
13.	<input type="checkbox"/>	An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.			
14.	<input checked="" type="checkbox"/>	A FIRST preliminary amendment.			
	<input type="checkbox"/>	A SECOND or SUBSEQUENT preliminary amendment.			
15.	<input type="checkbox"/>	A substitute specification.			
16.	<input type="checkbox"/>	A change of power of attorney and/or address letter.			
17.	<input checked="" type="checkbox"/>	Other items or information: Associate Power of Attorney			



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PATENT TRADEMARK OFFICE

JC05 Rec'd PCT/PTO 05 FEB 2001

U.S. APPLICATION NO. (if known) see 37 CFR 1.50 Unassigned 09/762248		INTERNATIONAL APPLICATION NO. PCT/FR99/01828		ATTORNEY'S DOCKET NUMBER 065691/0209	
18. <input checked="" type="checkbox"/> The following fees are submitted:				CALCULATIONS	
Basic National Fee (37 CFR 1.492(a)(1)-(5): Search Report has been prepared by the EPO or JPO.....\$860.00					
International preliminary examination fee paid to USPTO (37 CFR 1.482).....\$690.00					
No international preliminary examination fee paid to USPTO (37 CFR 1.482) but international search fee paid to USPTO (37 CFR 1.445(a)(2))\$710.00					
Neither international preliminary examination fee (37 CFR 1.482) nor International search fee (37 CFR 1.445(a)(2)) paid to USPTO \$1,000.00					
International preliminary examination fee paid to USPTO (37 CFR 1.482) and all claims satisfied provisions of PCT Article 33(2)-(4)\$100.00					
ENTER APPROPRIATE BASIC FEE AMOUNT =				\$860.00	
Surcharge of \$130.00 for furnishing the oath or declaration later than 20 Months from the earliest claimed priority date (37 CFR 1.492(e))					
Claims	Number Filed	Included in Basic Fee	Extra Claims	Rate	
Total Claims	6	- 20	= 0	x \$18.00	\$0.00
Independent Claims	2	- 3	= 0	x \$80.00	\$0.00
Multiple dependent claim(s) (if applicable)				\$270.00	
TOTAL OF ABOVE CALCULATIONS =				\$860.00	
Reduction by 1/2 for filing by small entity, if applicable.				\$0.00	
SUBTOTAL =				\$860.00	
Processing fee of \$130.00 for furnishing English translation later the 20 months from the earliest claimed priority date (37 CFR 1.492(f). +					
TOTAL NATIONAL FEE =				\$860.00	
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 per property +					
TOTAL FEES ENCLOSED =				\$860.00	
				Amount to be: refunded	\$
				charged	\$
<p>a. <input checked="" type="checkbox"/> A check in the amount of <u>\$860.00</u> to cover the above fees is enclosed.</p> <p>b. <input type="checkbox"/> Please charge my Deposit Account No. <u>19-0741</u> in the amount of \$0.00 to the above fees. A duplicate copy of this sheet is enclosed.</p> <p>c. <input checked="" type="checkbox"/> The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. <u>19-0741</u>. A duplicate copy of this sheet is enclosed.</p>					
<p>NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.</p>					
<p>SEND ALL CORRESPONDENCE TO:</p> <p>Foley & Lardner Washington Harbour 3000 K Street, N.W., Suite 500 Washington, D.C. 20007-5109</p>					
<p style="text-align: right;">  SIGNATURE NAME STEPHEN B. MAEBIUS REGISTRATION NUMBER 35,264 </p>					

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IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Attorney Docket No. 065691/0209

In re patent application of

Robert AMSON et al.

Serial No. Unassigned

Group Art Unit: Unassigned

Examiner: Unassigned

Filed: Concurrently

For: USE OF AN ANIMAL MODEL DEFICIENT IN P53 AND HAVING MEMORY DEFICIENCY AND/OR BEHAVIORAL DISORDERS FOR THERAPEUTIC PURPOSES

Preliminary Amendment

Commissioner for Patents
Washington, D.C. 20231

Sir:

Prior to action by the Examiner, please amend the captioned application as follows:

In the Claims:

Please cancel claims 1 to 10 and enter the following new claims:

11. (New) A method for identifying compounds with anti-anxiety activity, comprising:

(a) administering a test compound to an animal comprising at least one non-functional allele of the p53 gene, and

(b) determining whether said test compound decreases anxiety in said animal, in which test compounds that decrease anxiety in said animal are identified as compounds with anti-anxiety activity.

12. (New) The method of claim 11, wherein said animal is a mouse.

13. (New) The method of claim 11, wherein two alleles of the p53 gene are not functional.

14. (New) A method for identifying compounds with memory restoring activity, comprising:

(a) administering a test compound to an animal comprising at least one non-functional allele of the p53 gene, and

(b) determining whether said test compound restores memory in said animal, in which test compounds that restore memory in said animal are identified as compounds with memory restoring activity.

15. (New) The method of claim 14, wherein said animal is a mouse.

16. (New) The method of claim 14, wherein two alleles of the p53 gene are not functional.

002.444813.1

REMARKS

By the foregoing amendment, claims 11-16 are pending in the captioned application. An action on the merits is now awaited. Should there be any questions, the examiner is invited to contact the undersigned at the telephone number shown below.

Respectfully submitted,

Feb 1, 2001
Date

for Stephen B. Maebius 45,279
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002.444813.1

SPTS

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WO 00/07438

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USE OF AN ANIMAL MODEL DEFICIENT IN P53 AND HAVING
MEMORY DEFICIENCY AND/OR BEHAVIORAL DISORDERS FOR
THERAPEUTIC PURPOSES

5 The present invention relates to an animal model
having anxiety-related memory deficiency and/or behavioral
disorders. It also relates to the use of this animal model
for the screening and characterization of molecules capable
of acting on memory and/or anxiety. It also proposes the
10 use of a compound or a composition for the preparation of a
medicament intended for said treatments.

15 The inventors have indeed established a
relationship between a functionality defect in the
molecular pathway of the p53 gene, on the one hand, and
anxiety-related memory and/or behavioral disorders, on the
other hand.

20 While the role of the p53 gene as tumor suppressor
has been well established by a series of studies (Levine et
al., 1991; Eliyahu et al., 1989; Michalovitz et al., 1990;
Hollstein et al., 1991), the investigations on p53-
deficient mice which succumb rapidly to neoplasia have
contributed toward reinforcing this observation (Donchower
et al., 1992). Interestingly, at a specific level of their
development, a significant portion of these mice show major
25 impairments including the abnormal closure of the neural
tube (Sah et al., 1995; Armstrong et al., 1995) leading to
exencephaly and then anencephaly.

30 The inventors therefore focused on studying mice
having a deficiency in their p53 gene but born with an
unimpaired central nervous system (CNS). No previous study
has described any flagrant histopathological abnormality;
however, this does not exclude the possibility that the
nonfunctional p53 gene may influence, at a molecular level,
the transduction pathways giving rise to an abnormal
35 functioning of central nervous system.

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The inventors therefore focused on determining if mice homozygous or heterozygous with respect to the deficiency of their p53 gene showed any characteristic feature in their neurological and cognitive behavior.

5 Two groups of animals were studied, homozygous young mice p53-/- (that is to say in which the two alleles of the p53 gene are not functional) and heterozygous adult mice p53-/+ (in which only one of the two alleles of the p53 gene is not functional). This choice was dictated by
10 the fact that the homozygous mice p53-/- (or so-called "knock-out" mice) develop tumors at an early stage of their life while the heterozygous mice develop tumors much later. The experiments were therefore carried out over a period of time when the animal is not ill since mice carrying tumors
15 can have abnormal behavior which does not result from a neurological dysfunction but rather from the fact that they are suffering from their tumors. The mice deficient in their p53 gene and used in the context of the present invention are marketed by TACONIC FARMS, USA.

20 Two tests presented in detail below were carried out, namely the Morris swimming pool and the Open-Field.

The Morris swimming pool test (Morris et al., 1981, Morris et al., 1984), in which spatial learning and memory are evaluated, was used successfully in recent studies in
25 particular on model mice having symptoms of Alzheimer's disease (Nalbantoglu et al., 1997, Hsiao et al., 1996).

In the Open-Field test, behavioral parameters are measured (Archer et al., 1973, Walsh et al., 1976). This procedure has already been used in the context of the
30 evaluation of the psychological profile of mice having a behavior associated with fear (Gershenfeld et al., 1997) and to study the effects on behavior of the inactivated gene encoding the estrogen receptor (Ogawa et al., 1997).

35 These tests were carried out according to a "blind" design as regards the age and the genotype of the mice.

Figure 1 presents results obtained by the Morris swimming pool test. It presents the spatial bias index as a function of the genotype of the mice. Figures 1a and 1b relate to homozygous young mice p53^{-/-} and figures 1c and 1d relate to heterozygous adult mice p53^{+/-}. In both cases, the spatial bias index is calculated immediately after learning the procedural component of the test and then fifteen days later without additional training.

Figure 2 presents the results obtained by the Open-Field test consisting in counting the number of times the mouse crosses the central part of the field knowing that the anxiety which it can manifest is capable of preventing it from doing so. Figures 2a and 2c present the number of crossings of the central sector by the p53^{-/-} and p53^{+/-} mice, respectively, compared with the control mice. Figures 2b and 2d represent the percentage of the number of crossings of the central sector relative to the total number of crossings for the same groups of mice.

Figures 1e and 2e relate to the analysis of variance (ANOVA) by means of a diagram of repeat measurements, relating to the behavior of the mice. The square correlation (r^2) was calculated in order to explain the part of the variance resulting from the deficiency of the p53 gene.

Figure 3 also presents the results obtained by the Morris swimming pool test as a function of the sex of the mice. Once in the water, the mice have to learn to navigate toward the invisible platform. At the end of the learning, the mice are subjected to the spatial bias index test (F = female, M = male, WT = wild type). The two modes of variance analysis revealed a significant interaction of the genotypic sex ($F(1.43)=11.63$; $p=0.001$; $r^2=0.26$). Scheffé post-hoc test: Fp53^{-/-} versus FWT = 3.25; $p = 0.01$; Fp53^{-/-} versus Mp53^{-/-} = 2.77; $p = 0.043$.

Figure 4 illustrates the activation of p21Waf1 and

the repression of PS1 in the brain of adult mice p53-/- (this includes mice p53-/- that have while young performed the Morris swimming pool and Open-Field tests as reported above. Upon analysis, it was found that these mice had not yet developed tumors).

The Northern-blot analysis (a-f) of Waf1, PS1 and GAPDH (as control) is carried out on brains of young (a-c) and adult (d-f) mice. Lane 1 corresponds to the control mouse, lanes 2-3 to the male and female p53 knock-out mice. (a-c) correspond to the same Northern blot hybridized with different probes. No difference was noted in the expression of Waf1 and of PS1 between the control and the knock-out mice. For the adult mice, the hybridization is also carried out on the same Northern blot (d-f). The parentheses for (GAPDH) and (PS1) indicate what remains of the probes after dehybridization under nonstringent conditions. A high induction of p21 Waf1 (d) and a repression of PS1 (e) are noted in the p53-deficient mice.

The Western-blot analysis is carried out on protein extracts from the brains of the same adult animals (g-i), with anti-Waf1, anti-PS1 and anti-beta-tubulin (as control) antibodies. Lane 1 corresponds to the control mouse; lanes 2-3 to the p53 knock-out mice. An increased expression of p21Waf1 and a reduced expression of PS1 are observed in the p53-deficient mice.

Figure 5 illustrates apoptosis and the intracytoplasmic accumulation of amyloid-beta 42 in the brains of adult mice p53-/- as defined above.

(a) and (b) represent labeling of the isocortex with hematoxylin and with eosin in a control mouse and a knock-out mouse, respectively. The arrows indicate a thinning of the isocortex in the knock-out mouse (b). The magnification is 6.6 relative to the origin.

(c) and (d) represent staining by the TUNEL technique and the Harris hematoxylin counterstaining in the

control (c) and in the knock-out mouse (d). In (d), two positive nuclei are observed by the TUNEL technique (arrows). The magnification is 132.

5 (e) illustrates the quantification by the TUNEL technique.

(f-i) illustrate the immunohistochemical analysis using anti-amyloid-beta 42 specific antibodies. f and h correspond to the control mice, g and i are p53 knock-out mice which show an intracytoplasmic accumulation of amyloid-beta 42 (f-g: magnification $\times 40$, h-i magnification $\times 100$).

Results of the Morris swimming pool test

The young mice p53^{-/-} and the adult mice p53^{-/+} satisfy the test in a manner comparable to the control mice of the same age. This indicates that spatial learning in these mice is not affected by any deficiency in their p53 gene (Figure 1a and Figure 1c). In order to measure the memory of these mice, they were again subjected to the Morris swimming pool test after a period of two weeks and without additional training. The young mice p53^{-/-} have a lower spatial index but not significantly different from that shown by the control mice at the same age (Figure 1b). On the other hand, the adult mice p53^{-/+} have a significantly lower spatial index than that shown by the control mice (Figure 1d).

These results show that a memory deficiency is present in this group of animals since the real level of the spatial index, after a period of fifteen days, depends only on their capacity to recall the test. A histological examination of the brain of the adult mice p53^{-/+} was carried out but did not reveal any abnormality (data not shown). The fact that this memory deficiency is only visible in the adult mice p53^{-/+} and not in the young mice p53^{-/-} suggests that an age-dependent factor, that is to say the stage of maturation of the brain, may play a role

in the detection of memory deficiency. Although most studies of this type designed to detect behavioral or memory abnormalities, in neurodegenerative diseases, are carried out on adult mice, adult homozygous mice could not be studied because of the tumor development and that is the reason why the study was carried out on heterozygous mice.

Results of the Open-Field test

The young mice p53-/- (Figure 2a) and adult mice p53-/+ (Figure 2c) show a significantly lower number of crossings of the central sector compared with the control group of mice of the same age. Since the number of crossings of the central sector is proportional to the capacity of the mice to overcome their fear of moving away from the wall, in order to explore an unknown territory, these results suggest that the p53-deficient mice are more thigmotaxic than the corresponding normal mice (that is to say that they have a higher tendency to keep close to the walls). In order to exclude the possibility that a small number of crossings of the central sector may be due to a globally reduced locomotive activity, the number of crossings of the central sector was expressed as a percentage of the total number of crossings. This relative central activity is also significantly reduced in the two groups of p53-deficient mice (Figure 2b and Figure 2d).

The inventors then focused on going deeper into the results of the Morris swimming pool tests as reported above, obtained for the homozygous mice p53-/-, according to their sex. They therefore separated the measurements of the spatial index obtained for the male mice from those obtained for the female mice and compared them to the corresponding control group (Figure 3).

While the males p53-/- show no significant difference from the male and female wild-type controls, the females p53-/- show a performance which is significantly less than the control, this indicating lower capacities to

memorize and a poorer spatial learning.

The inventors then continued their investigations in order to determine if the disruptions in the functions of the central nervous system in the p53 knock-out mice were correlated with changes in the expression, in the brain, of p21Waf1 and PS1 since these two genes are regulated by wild-type p53 and are considered as modulators of apoptosis. They detected striking differences correlated with age. The young mice p53^{-/-} (two months old) have the same level of mRNA corresponding to p21Waf1 and PS1 as the control mice (Figure 4 a-c). By contrast, the adult mice p53^{-/-} (the same group on which the Morris swimming pool and Open-Field tests were carried out two months earlier) show a high increase in the expression of p21Waf1 and high repression of the expression of PS1 (Figure 4 d-f). Given the fact that PS1 undergoes proteolytic cleavage, the inventors tested its expression at the protein level. As shown in Figure 4h, the expression of the PS1 protein is very substantially reduced in the p53 knock-out mice. The inventors also tested the p53 knock-out mice in relation to mutations in the coding region of the cDNA for PS1. Nothing was found. These observations suggest that the older p53 knock-out mice overcompensate for the loss of p53 by over-expression of p21Waf with consequently a repression of PS1. Previous studies by the inventors showed that the increased expression of p21Waf1 resulted from the repression of PS1 with an induction of apoptosis and that the inhibition of the production of PS1 by an antisense cDNA induced apoptosis. This corroborates the notion that the reduction in the expression of PS1 in the brain could cause impairments in the functioning of the central nervous system through an increase in apoptosis.

The fact that young p53 knock-out mice already have deficiencies in relation to learning, memory and behavior before a measurable reduction in the expression of PS1 is

similar to what is found in neurodegenerative diseases. Abnormalities in behavior may be observed in patients who have been suffering from Alzheimer's disease for a long time before the terminal stage of the disease in which there is neurodegeneration. These data suggest a process which progresses slowly, in which the memory and behavioral abnormalities take place before a repression of PS1 becomes measurable.

The histopathological analyses combined with the TUNEL technique for evaluating apoptosis were performed on the same group of old animals showing a reduction in the expression of PS1 as indicated above. Three of the four mice showed no striking pathological abnormality. One knock-out mouse (female) showed a thinning of more than 50% of the isocortex with a very enlarged ventricle compared with the control animals (Figure 5a-b). The TUNEL experiment showed discrete apoptotic lesions (Figure 5d) in the brain without massive apoptosis. The quantification of the positive cells by TUNEL (Figure 5e) indicated that, whereas in the control brains only one case showed five positive cells out of 1740 counted, the mice p53-/- had up to 5.5% (25/465) and 13% (39/305) positive cells by TUNEL in the male and female animals, respectively (Figure 5e).

Because patients with Alzheimer's disease and transgenic mice with mutations in PS1 show neurodegeneration with apoptosis and accumulation of amyloid-beta 42, the inventors then tested for the presence of amyloid-beta 42 in the brains of the p53 knock-out mice. As shown in Figure 5h-i, the p53-deficient mice show high levels of amyloid-beta 42 in the cytoplasmic compartment.

In conclusion, these experiments indicate that the intact expression of p53 is necessary to ensure the complex functions of the central nervous system. These functions include memory and a standard behavior in anxiogenic situations. The precise mechanism by which the loss of the

functionality of p53 interferes with memory and behavior may be due to a deregulation of the p53 metabolic pathway at the molecular level.

Furthermore, it was shown that the mice p53-/- which showed deficiencies in memory, in their ability to learn, as well as an abnormal behavior had discrete apoptotic lesions in their brain. These impairments appear to indicate an age-dependent overcompensation of the p53 deficiency in the long term by an increase in the expression of p21Waf1 which leads to a repression of PS1. This in turn compromises the anti-apoptotic effect of PS1. These data indicate that the loss of the function of PS1 either through mutations as shown in early familial Alzheimer's diseases, or through the deregulation of expression as is the case in p53-deficient mice, activates the program of cell death. This is coupled with the impaired metabolism of APP (Amyloid Precursor Protein) leading to the accumulation of amyloid-beta 42. Thus, the inventors have demonstrated the basis for novel strategies for the treatment of neurodegenerative diseases.

Thus, surprisingly, the inventors have demonstrated that a mouse deficient in its p53 gene had learning and memory disorders and/or behavioral disorders such as anxiety.

Consequently, the present invention relates to an animal model, and in particular a mouse, having memory deficiency, the animal in question having at least one of the alleles of its p53 gene nonfunctional. This is the case with the mice p53-/+.

The model in question may in addition have behavioral disorders such as anxiety, and in this case, it is both alleles of the p53 gene in the animal which are not functional. This is the case with the homozygous mice p53-/-, but also as indicated above, the mice p53-/+ which also have behavioral disorders in an anxiogenic situation.

Thus, the animal model in question may be used for the screening of molecules capable of having an anxiolytic activity and/or of at least partially restoring memory.

Indeed, after having subjected mice p53-/+ to any learning test, it is known that two weeks later, they show a memory deficiency. They are then treated with a molecule (preferably suspected of acting on the memory), and the treated mice are again subjected to the test. The molecule will only be selected for additional investigations if it made it possible to significantly restore the memory of said mice.

In the context of tests conventionally carried out in this field, the animal model in accordance with the invention may also be used to determine the characteristics of a molecule which is known to be capable of acting on anxiety and/or memory, the characteristics relating to the pharmacodynamics, the pharmacokinetics and the toxicity of these molecules and the like.

Finally, the present invention also relates to the use of an agent that induces the metabolic pathway of the p53 gene, whether upstream or downstream thereof, for the preparation of a medicament intended for the treatment of memory and anxiety disorders and/or of neurodegenerative diseases. Indeed, the p53 gene is involved in a cascade of genes which interact with each other. The equilibrium of this cascade is characteristic of "a normal state" characterized by control of cell growth and of the process of apoptosis. It may therefore be advantageous to be able to act at different levels of the molecular pathway of the p53 gene according to the desired result.

Consequently, the present invention also relates to the case where said induction of the metabolic pathway of the p53 gene is an induction of the expression of a gene activated or inhibited by the p53 gene.

In particular, the abovementioned agent may in

addition exhibit an antitumor activity by activation of the metabolic or molecular pathway of the p53 gene. This agent may be for example the product of the expression of one of the genes involved in the cascade mentioned above.

5 The invention is not limited to the above description and the results obtained in the context of the abovementioned experiments will be understood more clearly by means of the presentation below of the two tests to which the mice were subjected.

10 METHODS: Behavioral measurements

The Morris swimming pool (Morris, 1981). The circular swimming pool made from PVC, of ivory color and 15 70 cm in diameter and 30 cm in height, is filled with water at $22 \pm 1^\circ\text{C}$, which is colored white by addition of the coloring Opacifier 631®, to a height of 12 cm below the edge of the wall. A circular platform 5 cm in diameter serving as the objective is immersed 0.5 cm under the 20 water, 7 cm away from the wall. The device is placed in a rectangular room. Two rectangular markers (50 x 30 cm), one black and one with black and white stripes, are attached to two adjacent walls 1.5 meters away from the swimming pool. A video camera, placed vertically above the swimming pool 25 makes it possible to record the movements of the mice on a video tape in order to analyze them subsequently. The experimentalist is hidden behind a white curtain. Placed in the water, facing the wall, in a quadrant that varies in a pseudorandom manner from one test to another, the mice have 30 to learn to navigate toward the platform using the distal visual markers available in the room. After a practice session of three trials intended for learning the procedural component of the test, the mice are subjected to three consecutive trials per day for 4 days. The maximum 35 time allowed to reach the platform was set at 60 seconds.

If a mouse cannot reach the platform within this period of time, it is guided to it by the experimentalist. The sum of the three lag times constitutes the score for the session. After the last trial of the third session, the mouse is subjected to a spatial bias test intended to check the accuracy of the spatial learning (short-term memory). The platform is then removed from the device and the mouse, starting from the opposite quadrant, has to swim in search of it for one minute. The route is recorded on a videotape and a spatial bias index is then calculated. It corresponds to the difference between the number of times it crosses an 8 cm ring surrounding the position of the platform and the mean number of times it crosses the three annuli placed symmetrically in the other three quadrants. Another spatial bias test is carried out after a period of 15 days without additional training (long-term memory).

Open-field (Archer, 1973; Walsh and Cummins, 1976).

The mice are introduced individually into a grey PVC cylinder 40 cm in diameter and 30 cm in height, placed on a sheet of white paper through which it is possible to distinguish the lines which divide the floor of the open-field into 7 sectors having the same surface area (1 central sector and 6 peripheral sectors). This device, illuminated with white light diffusing through frosted glass (125 lux) constitutes for the mice a moderately anxiogenic situation. The mice are subjected to three daily sessions of 15 minutes, 45 minutes apart, this being for 4 consecutive days. The behavioral measurements are performed for the first 5 minutes of each session. They consist essentially in counting the number of times the mice "are bold enough" to move away from the side walls of the cylinder in order to cross the central sector of the Open-Field.

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CLAIMS

1. Use of an animal model showing memory deficiency and/or behavioral disorders such as anxiety, in which at least one of the alleles of the p53 gene is not functional, for the screening of molecules capable of exhibiting anxiolytic activity.

2. Use of an animal model as defined in claim 1, for determining the characteristics of a molecule having an anxiolytic activity such as the pharmacodynamics, the pharmacokinetics and the toxicity.

3. Use of an animal model according to claim 1 or 2, characterized in that the two alleles of the p53 gene are not functional.

4. Use of an animal model as defined in claim 1, for the screening of molecules capable of at least partially restoring memory.

5. Use of an animal model as defined in claim 1, for determining the characteristics of a molecule capable of at least partially restoring memory such as the pharmacodynamics, the pharmacokinetics and the toxicity.

6. Use of an animal model according to one of claims 1 to 5, characterized in that the animal is a mouse.

7. Use of an agent for inducing the metabolic pathway of the p53 gene, upstream or downstream of said gene, for the preparation of the medicament intended for the treatment of memory disorders, anxiety disorders and/or neurodegenerative diseases.

8. Use of an agent according to claim 7 in which the induction of the metabolic pathway of the p53 gene is an induction of the expression of a gene activated by the p53 gene.

9. Use of an agent according to claim 7, in which the induction of the metabolic pathway of the p53 gene is an induction of the expression of a gene inhibited by the

p53 gene.

10. Use of an agent according to one of claims 7 to 9, characterized in that the said agent has furthermore an anti-tumor activity through the activation of the metabolic pathway of the p53 gene.

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FIGURE 1

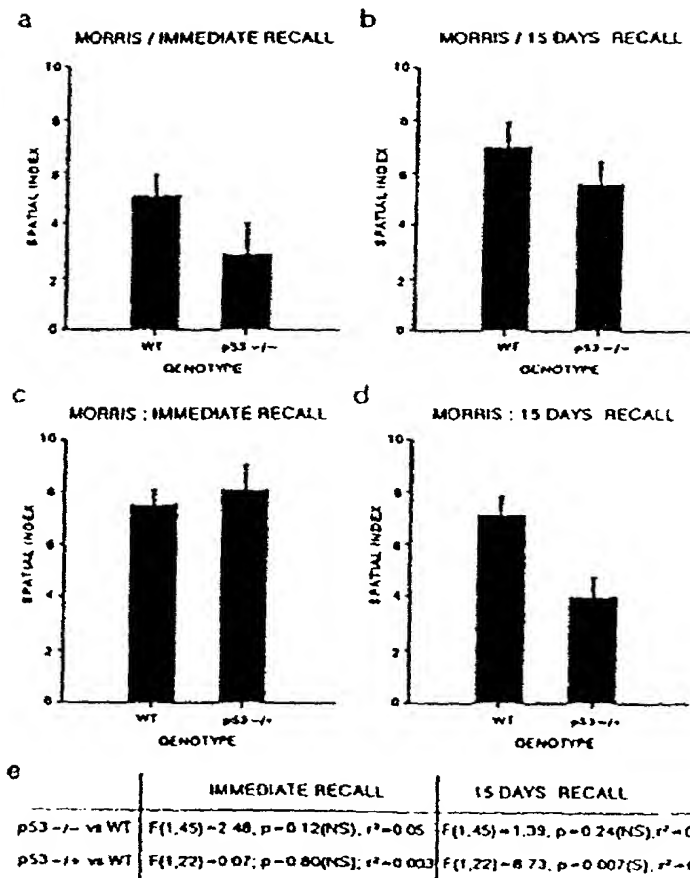


FIGURE 2

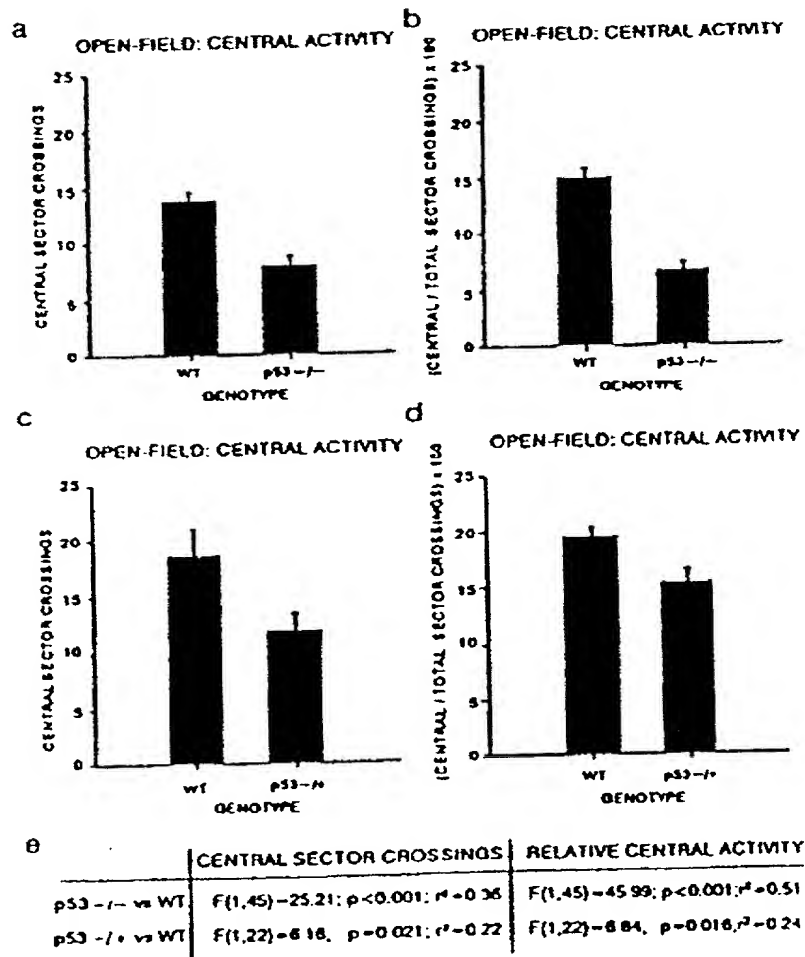
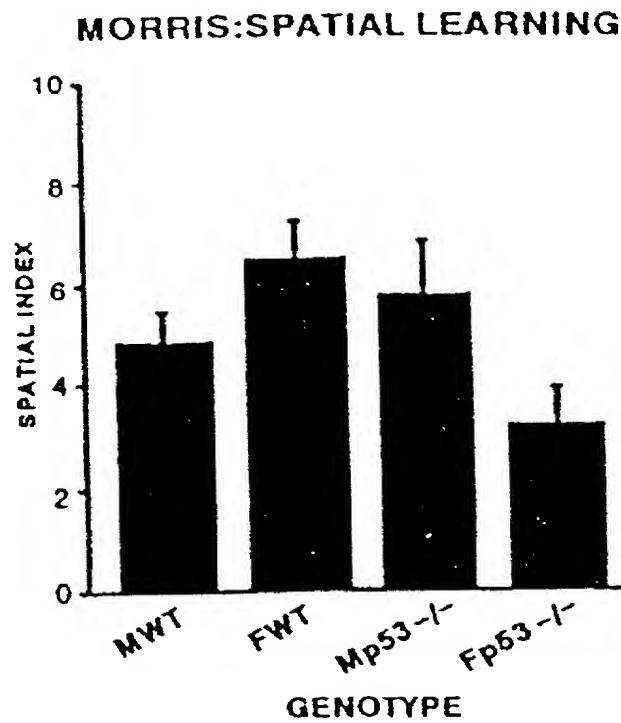


FIGURE 3



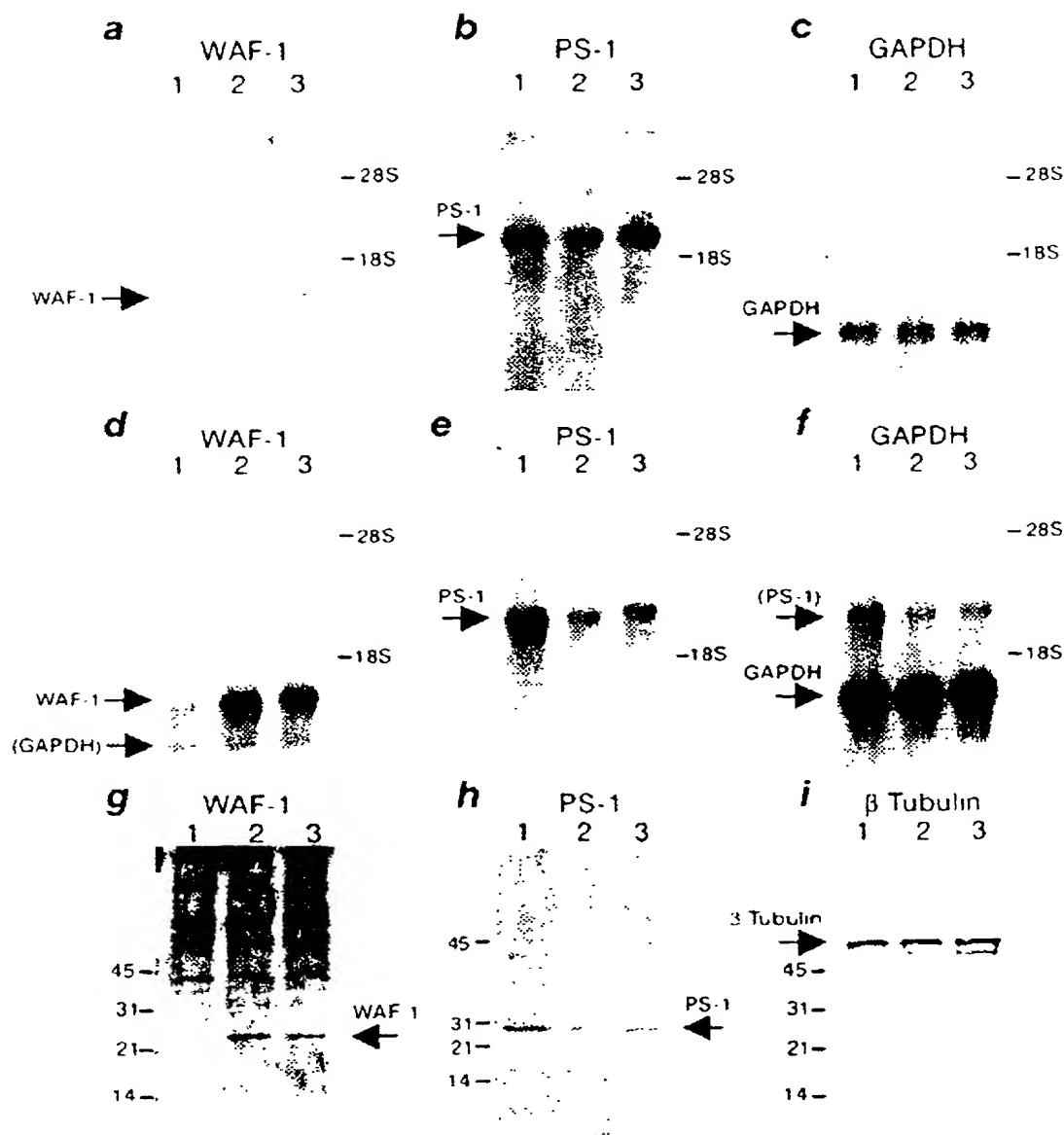
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FIGURE 4



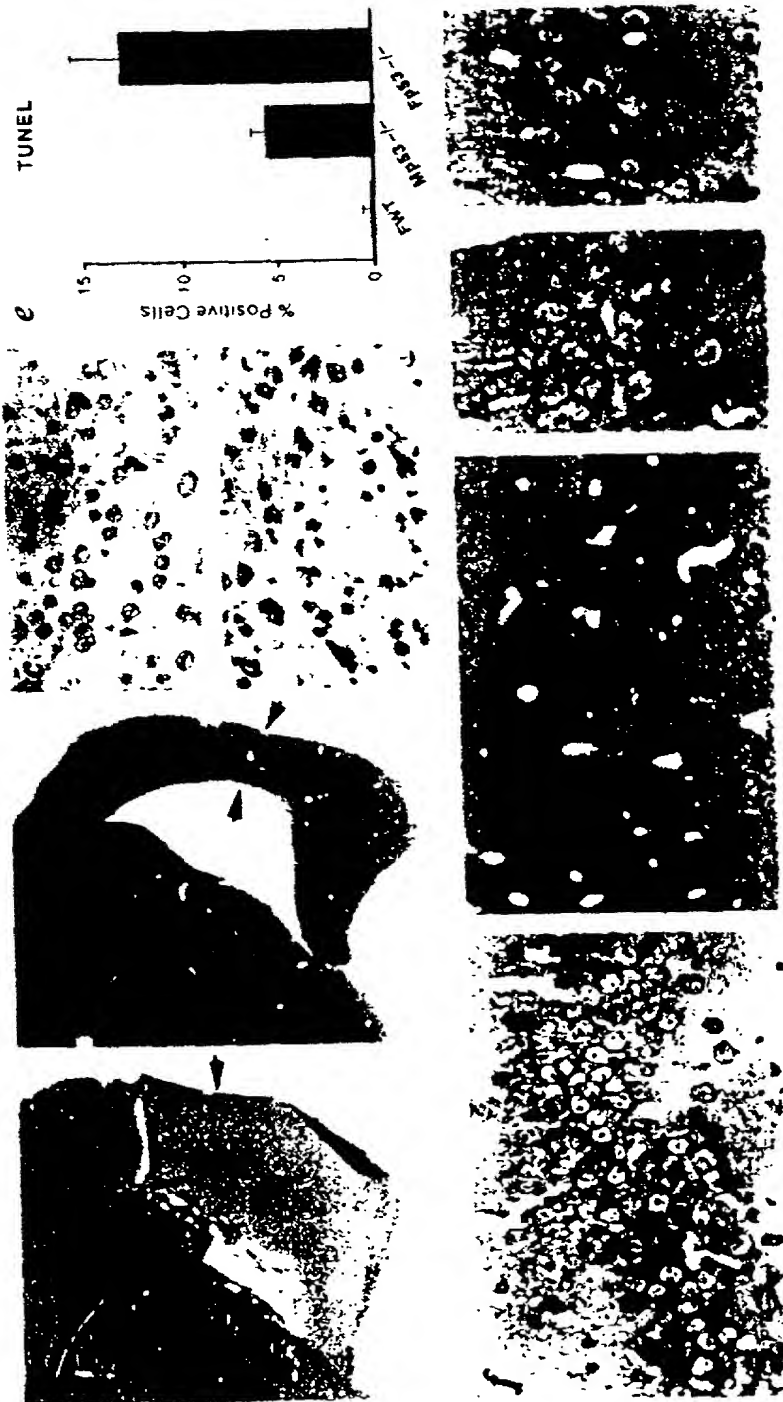
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FIGURE 5



IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Attorney Docket No. 065691/0209

In re patent application of

Robert AMSON et al.

Serial No. Unassigned

Group Art Unit: Unassigned

Examiner: Unassigned

Filed: Concurrently

For: USE OF AN ANIMAL MODEL DEFICIENT IN P53 AND HAVING MEMORY DEFICIENCY AND/OR BEHAVIORAL DISORDERS FOR THERAPEUTIC PURPOSES

ASSOCIATE POWER OF ATTORNEY

Assistant Commissioner for Patents
Washington, D.C. 20231

Sir:

The undersigned attorney of record hereby appoints Stephen B. Maebius, Registration No. 35,264 as an associate attorney with full power of association, substitution and revocation, to prosecute the above-identified application and transact all business in the Patent and Trademark Office connected therewith.

Respectfully submitted,



February 5, 2001

Date

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DECLARATION AND POWER OF ATTORNEY

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name.

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

the specification of which is attached hereto unless the following box is checked:

☐ was filed on July 26, 1999 as United States Application Number or PCT International Application Number FR99/01 828 and was amended on _____ (if applicable).

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is known by me to be material to patentability as defined in Title 37, Code of Federal Regulations § 1.56.

I hereby claim foreign priority benefits under Title 35, United States Code, § 119(a)-(d) or § 365(b) of any foreign application(s) for patent or inventor's certificate, or § 365(a) of any PCT International application which designated at least one country other than the United States, listed below and have also identified below any foreign application for patent or inventor's certificate, or PCT International application having a filing date before that of the application on which priority is claimed:

PRIOR FOREIGN APPLICATION(S)

NUMBER	COUNTRY	DAY/MONTH/YEAR FILED	PRIORITY CLAIMED
98 10076	FRANCE	August 5, 1998	Y

I hereby claim the benefit under Title 35, United States Code § 119(e) of any United States provisional application(s) listed below.

APPLICATION NO.	FILING DATE

I hereby claim the benefit under Title 35, United States Code, § 120 of any United States application(s), or § 365(c) of any PCT International application designating the United States, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT International application in the manner provided by the first paragraph of Title 35, United States Code, § 112, I acknowledge the duty to disclose information which is known by me to be material to patentability as defined in Title 37, Code of Federal Regulations § 1.56 which became available between the filing date of the prior application and the national or PCT International filing date of this application:

APPLICATION SERIAL NO.	FILING DATE	STATUS: PATENTED, PENDING, ABANDONED

I hereby appoint as my attorneys, with full powers of substitution and revocation, to prosecute this application and transact all business in the Patent and Trademark Office connected therewith: Stephen A. Bent, Reg. No. 29,768; David A. Blumenthal, Reg. No. 26,257; William T. Ellis, Reg. No. 26,874; John J. Feldhaus, Reg. No. 28,822; Patricia D. Granados, Reg. No. 33,683; John P. Isacson, Reg. No. 33,715; Donald D. Jeffery, Reg. No. 19,980; Eugene M. Lee, Reg. No. 32,039; Richard Linn, Reg. No. 25,144; Peter G. Mack, Reg. No. 26,001; Brian J. McNamara, Reg. No. 32,789; Sybil Meloy, Reg. No. 22,749; George E. Quillin, Reg. No. 32,792; Colin G. Sandercock, Reg. No. 31,298; Bernhard D. Saxe, Reg. No. 28,665; Charles F. Schiff, Reg. No. 27,590; Richard L. Schwaab, Reg. No. 25,479; Arthur Schwartz, Reg. No. 22,115; Harold C. Wegner, Reg. No. 25,258.

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